

# resDetect™ Human Interleukin-7 (IL-7) ELISA Kit (Residue Testing)

Catalog Number: RES-A055

Pack Size: 96 tests

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedure



## **INTENDED USE**

Human Interleukin-7 (IL-7) ELISA Kit (Residue Testing) was developed for the detection and quantitative determination of GMP human IL-7 in samples from CAR-T product preparation processing. It is intended for research use only (RUO).

# **BACKGROUND**

Interleukin-7 (IL-7) is a pleiotropic cytokine of the chemokine family and has broad immune effects. IL-7 stimulates the differentiation of hematopoietic stem cells into lymphoid progenitor cells, and can also stimulate the proliferation of all lymphocytes (B cells, T cells, and NK cells).

To support the development of CAR-T drugs, ACROBiosystems independently developed human Interleukin-7 (IL-7) ELISA Residue Testing kit via rigorous methodological validation, which is used for detection of GMP human IL-7 in samples from CAR-T product preparation processing for evaluation the quality of CAR-T products in drug development and CMC quality control stages.

# PRINCIPLE OF THE ASSAY

This assay kit is used to measure the levels of human Interleukin-7 (IL-7) by employing a standard sandwich-ELISA format. The micro-plate in the kit has been pre-coated with Anti-IL-7 Antibody. Firstly, add the standard samples provided in kit and your samples to the plate, incubate and wash the wells. Then add the Biotin-Anti-IL-7 Antibody to the plate and form Antibody-antigen-biotinylated antibody complex, incubate and wash the wells. Next add Streptavidin-HRP to the plate, incubate and wash the wells. At last, load the substrate into the wells and monitor solution color from blue to yellow. The reaction is stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value reflects the amount of human IL-7 bound.

#### **PRECAUTIONS**

1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.



- 2. The kit is suitable for cell supernatant and serum and plasma samples.
- 3. Do not use reagents past their expiration date.
- 4. Do not mix or substitute reagents with those from other kits or other lot number kits.
- 5. If samples generate values higher than the highest standard, dilute the samples with the appropriate calibrator diluent and repeat the assay. If cell supernatant samples need step dilution, except for the final dilution with diluent, other intermediate dilutions can be in cell culture medium.
- 6. Differences in test results can be caused by a variety of factors, including laboratory operator, pipette usage, plate washing technique, reaction time or temperature, and kit storage.
- 7. This kit is designed to remove or reduce some endogenous interference factors in biological samples, and not all possible influencing factors have been removed.

# **MATERIALS PROVIDED**

Table 1. Materials provided

				Stor	rage
Catalog	Components	Size (96 tests)	Format	Unopened	Opened
RES055-C01	Pre-coated Anti-IL-7 Antibody Microplate	1 plate	Solid	2-8°C	2-8°C
RES055-C02	Human IL-7 Standard	20 μg	Power	2-8°C	-70°C
RES055-C03	Biotin-Anti-IL-7 Antibody	20 μg	Power	2-8°C	-70°C
RES055-C04	Streptavidin-HRP	50 μL	Liquid	2-8°C, avoid light	2-8°C, avoid light
RES055-C05	10×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C
RES055-C06	2×Dilution Buffer	50 mL	Liquid	2-8°C	2-8°C
RES055-C07	55-C07 Substrate Solution		Liquid	2-8°C, avoid light	2-8°C, avoid light
RES055-C08	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

# **SRORAGE**

- 1. Unopened kit should be stored at 2°C -8°C upon receiving.
- 2. The opened kit should be stored per Table 1. The shelf life is 30 days from the date of opening.

*Note:* a. Do not use reagents past their expiration date.

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b. Find the expiration date on the outside packaging.

## REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or multi-channel micropipettes and pipette tips: need to meet  $10~\mu L$ ,  $300~\mu L$ ,  $1000~\mu L$  injection requirements;

37°C Incubator;

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Tubes: 1.5 mL,10 mL;

Timer:

Reagent bottle;

Deionized or distilled water.

## REAGENT PREPARATION

Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, place the sample in an 37°C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

According to Table 2, prepare the provided lyophilized product into a storage solution with ultrapure water, dissolve at room temperature for 15 to 30 minutes, and mix by gently pipetting, avoiding vigorous shaking or vertexing. The reconstituted storage solution should be stored at  $-70^{\circ}$ C. It is recommended that the number of freezing and thawing should not exceed 1 time, the size of the aliquot should not be less than 5  $\mu$ g.

**Note:** Considering inevitable minor quantitation variations between protein batches, it is also reasonable to generate the standard curve with specific lot of proteins used for current production for even better accuracy.

**Table 2. Preparation method** 

ID	Components Size (96 T)		Storage solution concentration.	Reconstituted water Vol.
RES055-C02	Human IL-7 Standard	20 μg	200 μg/mL	100 μL
RES055-C03	Biotin-Anti-IL-7 Antibody	20 μg	200 μg/mL	100 μL

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# **RECOMMENDED SAMPLE PREPARATION**

## 1. Working Solution Preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

1.2 Preparation of 1×Dilution Buffer:

Dilute 50 mL 2×Dilution Buffer with 1×Washing Buffer to 100 mL.

1.3 Preparation of Biotin-Anti-IL-7 Antibody working fluid:

Dilute Biotin-Anti-IL-7 Antibody to 0.1  $\mu$ g/mL with 1×Dilution Buffer. Please prepare it for one-time use only.

1.4 Preparation of Streptavidin-HRP working fluid:

Dilute Streptavidin-HRP at 1:2000 with 1×Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

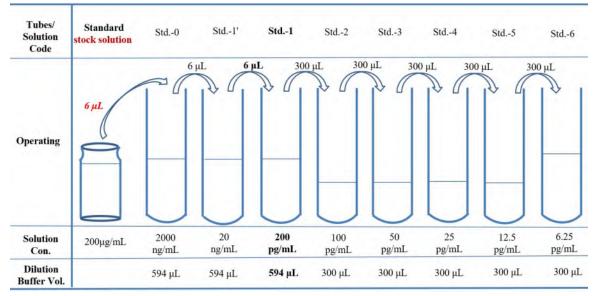
- 1.5 Sample preparation
- a. If the sample to be tested is the serum or plasma, dilute test sample at 1:5 with 1×Dilution Buffer. The volume ratio of sample to diluent is 1:4.
- b. If the sample to be tested is the cell supernatant, dilute test sample at 1:2 with 1×Dilution Buffer. The volume ratio of sample to diluent is 1:1.

#### 2. Preparation of Standard curve

The concentration of the reconstituted Human IL-7 Calibrator (RES055-C02) is 200  $\mu$ g/mL, prepare (Std.-0) by diluting 6  $\mu$ L the reconstituted Human IL-7 Calibrator into 594  $\mu$ L Sample Dilution Buffer, mix gently well. Then prepare Std.-1' by diluting 6  $\mu$ L Std.-0 into 594  $\mu$ L Sample Dilution Buffer. At last, prepare the highest concentration of standard curve, **Std.-1** (200 pg/mL), by diluting 6  $\mu$ L Std.-1' into 594  $\mu$ L Sample Dilution Buffer. Prepare 1:1 serial dilution for the standard curve as follows: Pipette 300  $\mu$ L of Sample Dilution Buffer into each tube. Make sure to mix well every time. Sample Dilution Buffer serves as blank.

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#### 3. Add Samples

Add 100  $\mu$ L Calibrator and samples to each well. For blank Control wells, please add 100  $\mu$ L Dilution Buffer.

*Note*: It is recommended to set double holes for samples and standard curves to be tested.

#### 4. Incubation

Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

#### 5. Washing

Remove the remaining solution by aspiration, add 300  $\mu$ L of 1×Washing Buffer to each well, soak for 10 s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

#### 6. Add Biotin-Anti-IL-7 Antibody

For all wells, add 100  $\mu$ L Biotin- Anti-IL-7 Antibody (dilute to 0.1  $\mu$ g/mL) working solution. Please prepare it for one-time use only.

#### 7. Incubation

Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

#### 8. Washing

Repeat step 5.

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### 9. Add Streptavidin-HRP

For all wells, add 100 µL Streptavidin-HRP (dilute at 1:2000) working solution. Please prepare it for one-time use only, avoid light.

#### 10. Incubation

Seal the plate with microplate sealing film and incubate at room temperature for 30 min.

## 11. Washing

Repeat step 5.

#### 12. Substrate Reaction

Add 100 µL Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at room temperature for 20 min, avoid light.

#### 13. Termination

Add 50 µL Stop Solution to each well and tap the plate gently to allow thorough mixing.

Note: The color in the wells should change from blue to yellow.

#### 14. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 10 minutes.

**Note**: To reduce the background noise, subtract the value read at  $OD_{450nm}$  with the value read at  $OD_{630nm}$ .

# **CALCULATION OF RESULTS**

- 1. Calculate the mean absorbance for each standard, control and sample and subtract average zero standard optical density (OD).
- 2. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.
- 3. Normal range of Standard curve:  $R^2 \ge 0.9900$ .
- 4. Detection range: 6.25 pg/mL-200 pg/mL. If the OD value of the sample to be tested is higher than 200 pg/mL, the sample shall be diluted with dilution buffer and assay repeated. If the OD value of the sample to be tested is lower than 6.25 pg/mL, the sample should be reported.

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# **QUICK GUILD**

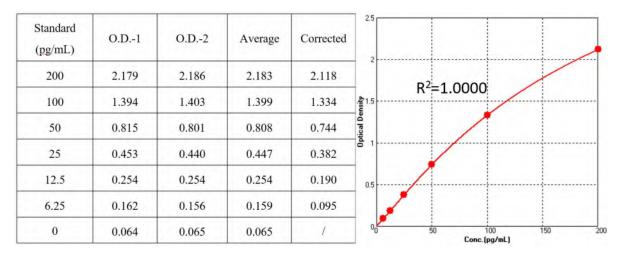


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# **TYPICAL DATA**

For each experiment, a standard curve needs to be set for each micro-plate, and the specific OD value may vary depending on different laboratories, testers, or equipments. The following example data is for reference only. The sample concentration was calculated based on the results of the standard curve.



## **SENSITIVITY**

The minimum detectable concentration of human IL-7 is 2.148 pg/mL. The minimum detectable concentration was determined by adding twice standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

## **PRECISION**

#### 1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

#### 2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

	In	tra-assay Precisi	on	In	ter-assay Precisi	on
Sample	1	2	3	1	2	3
n	10	10	10	3	3	3

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RES55-EN.01

Mean (pg/mL)	142.027	58.304	14.569	147.134	59.601	15.043
SD	6.094	1.994	0.477	4.785	1.475	0.417
CV (%)	4.3%	3.4%	3.3%	3.3%	2.5%	2.8%

*Note*: The example data is for reference only.

# **RECOVERY**

Three parts of blank serum were added with different concentrations of human IL-7, and the serum without human IL-7 was used as background to calculate the recovery rate. The range of the recovery rate is 92.5-102.4%, and the average recovery is 98.8%.

Sample Type	Average % Recovery	Range
Serum(n=5)	98.8%	92.5-102.4%

# **LINEARITY**

To assess the linearity of the assay, samples spiked with high concentrations of human IL-7 were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)	Serum
1:2	Average Recovery (%)	99.7	100.3	99.9
1:2	Range (%)	97.5-101.3	97.5-103.7	97.5-101.9
1.4	Average Recovery (%)	101.0	100.6	100.1
1:4	Range (%)	97.5-104.3	98.4-103.1	97.6-102.5
1:8	Average Recovery (%)	106.4	99.8	107.4
1:8	Range (%)	102.0-108.9	97.1-103.2	103.6-109.3
1:16	Average Recovery (%)	107.9	99.1	117.0
1.10	Range (%)	105.6-112.8	98.7-99.5	115.0-119.3

*Note*: The example data is for reference only.

# **SPECIFICITY**

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This assay recognizes natural and recombinant human IL-7. No cross-reactivity was observed when this kit was used to analyze the following recombinant factors.

Human							
TNF-alpha	Anti-CD28	IL-18	IL-2	Flt-3 Ligand			
GM-CSF	Anti-CD137	IL-17A	IL-15	Anti-CD3			
G-CSF	IL-4	L1R	Thrombopoietin-TPO	SCF			
M-CSF	IL-12B	VEGF165	IL-3	BMP-2			
IL-6	IL-10	IL-5	IL-11	FGF basic			
IFN-gamma	IL-21	IL-8	TGF-beta 1				

# **INTERFERING SUBSTANCES**

Verify potential matrix effects by adding different levels of DMSO and HSA to the diluted buffer.

Additive	Tolerated concentration
DMSO	20%
HSA	5%

# **CALIBRATION**

This immunoassay is calibrated against a highly purified *E. coli*-expressed recombinant human IL-7 (90/530). Reference Reagent is calibrated by NIBSC/WHO in April 2013.

NIBSC/WHO (90/530) approximate value (U/mL) =  $0.520 \times \text{Human IL-7 value (pg/mL)}$ 

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# **PLATE LAYOUT**

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Std1	Std1	()				()	()	()	()		()
В	Std2	Std2	(	$(\underline{\cdot\cdot})$	()		()	$(\cdots)$	()	$(\cdots)$		()
С	Std3	Std3	$\left( \begin{array}{c} \cdots \end{array} \right)$	$(\cdots)$	()		()	$(\cdots)$	()	$( \dots )$		()
D	Std4	Std4	$\left( \begin{array}{c} \cdots \end{array} \right)$	$(\cdots)$	)		()	$(\cdots)$	()	$(\cdots)$	( <u></u> )	()
E	Std5	Std5	( $$ $)$	(	)		()	$\left( \cdots \right)$	()	$( \dots )$	()	()
F	Std6	Std6	( $$ $)$	$(\cdots)$			()	( $$ $)$	()	$( \dots )$	()	()
G	Blank	Blank	(	$( \cdots )$			()	( $$ $)$	()	$( \dots )$		$\left( \begin{array}{c} \cdots \end{array} \right)$
Н	Blank	Blank	( )	$( \dots )$	( )	()	( )	$( \dots )$	( )	( )	( )	( )

Note: Blank is a Blank Dilution Buffer hole.

# **TROUBLESHOOTING GUIDE**

Problem	Cause	Solution	
Poor standard curve	* Inaccurate pipetting	* Check pipettes	
Lauga CV	* Inaccurate pipetting	* Check pipettes	
Large CV	* Air bubbles in wells	* Remove bubbles in wells	
High healtonound	* Plate is insufficiently washed	* Review the manual for proper wash.	
High background	* Contaminated wash buffer	* Make fresh wash buffer	
Very low readings across	* Incorrect wavelengths	* Check filters/reader	
the plate	* Insufficient development time	* Increase development time	
Samples are reading too			
high, but standard curve	* Samples contain cytokine	* Dilute samples and run again	
looks fine	levels above assay range		

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		* Assay set-up should be continuous - have all standards and samples prepared appropriately
Drift	* Interrupted assay set-up  * Reagents not at room temperature	before commencement of theassay  * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts

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